

## New Zealand Society of Animal Production online archive

This paper is from the New Zealand Society for Animal Production online archive. NZSAP holds a regular annual conference in June or July each year for the presentation of technical and applied topics in animal production. NZSAP plays an important role as a forum fostering research in all areas of animal production including production systems, nutrition, meat science, animal welfare, wool science, animal breeding and genetics.

An invitation is extended to all those involved in the field of animal production to apply for membership of the New Zealand Society of Animal Production at our website [www.nzsap.org.nz](http://www.nzsap.org.nz)

[View All Proceedings](#)

[Next Conference](#)

[Join NZSAP](#)

The New Zealand Society of Animal Production in publishing the conference proceedings is engaged in disseminating information, not rendering professional advice or services. The views expressed herein do not necessarily represent the views of the New Zealand Society of Animal Production and the New Zealand Society of Animal Production expressly disclaims any form of liability with respect to anything done or omitted to be done in reliance upon the contents of these proceedings.

This work is licensed under a [Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License](http://creativecommons.org/licenses/by-nc-nd/4.0/).



You are free to:

**Share**— copy and redistribute the material in any medium or format

Under the following terms:

**Attribution** — You must give [appropriate credit](#), provide a link to the license, and [indicate if changes were made](#). You may do so in any reasonable manner, but not in any way that suggests the licensor endorses you or your use.

**NonCommercial** — You may not use the material for [commercial purposes](#).

**NoDerivatives** — If you [remix, transform, or build upon](#) the material, you may not distribute the modified material.

<http://creativecommons.org.nz/licences/licences-explained/>

## BRIEF COMMUNICATION: Measurement of sole thickness and distance to distal phalanx using a portable ultrasound machine

L.J. LAVEN\*, J.K. MARGERISON, R.A. LAVEN

Institute of Veterinary, Animal and Biomedical Sciences, Massey University, Private Bag 11-222, Palmerston North 4442, New Zealand

\*Corresponding author: l.j.laven@massey.ac.nz

**Keywords** cattle; ultrasound; claw; sole thickness; distal phalanx.

### INTRODUCTION

Using a high specification ultrasound machine, with the ultrasound transducer applied to the sole surface, Kofler *et al.* (1999) were able to estimate sole and soft tissue thickness in claws from bovine cadavers. These ultrasound estimates were significantly correlated with measurements, made using mechanical callipers, of the same parameters in claws after sectioning. Moreover, Kofler *et al.* (1999) demonstrated that it was possible to image the internal structures of the bovine claw in the live animal and also reported the use of a portable ultrasound. They did however state that the images were of lower quality, due to the poorer resolution of the transducer, and were more difficult to obtain. In contrast, van Amstel *et al.* (2003, 2004a, 2004b) were able to visualise the sole and corium using a portable ultrasound, but did not assess the accuracy of the measurements in cadaver claws. As a consequence, this paper aims to assess bovine claws collected from cattle post mortem, to measure claw sole thickness and distance to distal phalanx using a portable ultrasound machine and to compare these with measurement collected using electronic callipers following sectioning of the frozen claws, such that changes in sole thickness could be assessed in future longitudinal studies with lactating dairy heifers.

### MATERIALS AND METHODS

The distal limbs were collected from 24 dairy cows, eight had died within the previous 24 hours of conditions unrelated to lameness, three were culled dairy cattle euthanased on-farm and the remainder were collected from a local abattoir. The age, breed and cause of death of each animal were recorded. On removal, each limb was identified using an elastic band of a specified colour. After collection, the limbs were washed and a latex glove placed over the sawn end of the limb to prevent moisture loss. All four limbs from each cow were placed in an individual plastic bag, sealed with a cable tie, and stored at 3 to 5°C, prior to ultrasound assessment. All ultrasound measurements were completed within 72 hours post mortem.

### Ultrasound assessment of claws

The ultrasound machine used for this study was a Mindray DP 6600 (Mindray, Szechuan, China). Prior to examination the solar surface of each claw was lightly pared. The transducer was protected by a vinyl glove containing acoustic coupling gel prior to use and additional gel was then applied to the claw surface to aid coupling between the claw surface and transducer. The transducer was placed along a line perpendicular to and bisecting the line from the abaxial groove to the axial border, which was also perpendicular to, and bisecting, the line from the end of the axial white line to the abaxial border (modified from Råber *et al.*, 2004). The ultrasound measurements were taken at two sites; the tip of the distal phalanx (Site 1) and 25 mm towards the heel (Site 2). All measurements were completed with the probe set at a frequency of 5 MHz. Two measurements were taken at each site; the distance to the distal phalanx from the external sole surface (DP) and the distance from the external sole surface to the internal sole surface (STh). The internal sole surface was visualised as a thin hyperechoic line of a continuous or interrupted nature above an anechoic region (Figure 1).

### Calliper measurement of distance to distal phalanx and sole thickness

The limbs were frozen at –20°C, directly following ultrasonography, for at least 24 hours before being sectioned. The frozen claws were sectioned sagittally along the line where the transducer was placed, using a band saw. DP and STh were measured using electronic callipers at Sites 1 and 2.

### Statistical analysis

Pearson's correlation was used to assess the relationship within all the parameters measured using the electronic callipers, and within the four ultrasound measurements, and to evaluate the association between the measurements obtained using ultrasound and measurements obtained using electronic callipers. All statistical analysis was completed using SPSS 16, (SPSS Inc., Chicago, Illinois, USA).



reported to interfere with imaging Kofler *et al.* (1999) and van Amstel *et al.* (2003, 2004a). While Kofler *et al.* (1999) successfully imaged the internal sole surface, as a thin hyper echoic line, in 86% (86/100) of the claws; they reported that the line was either interrupted or incomplete in only 9% (8/86) of the claws. This interruption was a more frequent finding of 14% (13/90) in this study.

In this study minimal claw preparation was undertaken, prior to obtaining ultrasound images, so as to not alter sole thickness or cause greater sole thinning. Despite this, good surface contact was achieved, most likely due to the relatively smooth and non-flaky nature of the sole horn. This was increased by the placement of the transducer probe into a glove filled with acoustic ultrasound gel, which moulded to the contour of the sole.

The present study used two measurements along a repeatable line, parallel to the long axis of the claw. This line was employed to allow collection of histological specimens for light microscopy from standard sites (Räber *et al.*, 2004). This proved to be a practical method for probe application and landmark recognition.

The results in this study agree with those reported by Kofler *et al.* (1999) in that correlations obtained for the distance to distal phalanx between ultrasound and calliper measurements were high for all sites measured. However, this study resulted in moderate correlations between sole thickness at Site 1 and no significant correlation at Site 2. Kofler *et al.* (1999) reported high correlations levels for these sites. Thus, it was concluded that, applying the methods used in this study, a Mindray DP 6600 portable ultrasound could be used to accurately estimate DP, however it was not considered sufficiently accurate for the direct estimation of STh.

## ACKNOWLEDGEMENTS

Financial support from the Department of Comparative Physiology and Anatomy, Institute of Food, Nutrition and Human Health at Massey University and McGeorge Research Fund. R. Neil Ward, Mike Hogan and Kim Fraser provided technical support.

## REFERENCES

- Kofler, J.; Kubber, P.; Henninger, W. 1999: Ultrasonographic imaging and thickness measurement of the sole horn and the underlying soft tissue layer in bovine claws. *The Veterinary Journal* **157**: 322-31.
- Räber, M.; Lischer, C.J.; Geyer, H.; Ossent, P. 2004: The bovine digital cushion - a descriptive anatomical study. *The Veterinary Journal* **167**: 258-64.
- van Amstel, S.R.; Palin, F.L.; Rorhbach, B.W.; Shearer, J.K. 2003: Ultrasound measurement of sole horn thickness in trimmed claws of dairy cows. *Journal of American Veterinary Medical Association* **223**(4): 492-494.
- van Amstel, S.R.; Palin, F.L.; Shearer, J.K. 2004a: Measurement of the thickness of the corium and subcutaneous tissue of the hind claws of dairy cattle by ultrasound. *The Veterinary Record* **155**: 630-633.
- van Amstel, S.R.; Shearer, J.K.; Palin, F.L. 2004b: Moisture content, thickness, and lesions of sole horn associated with thin soles in dairy cattle. *Journal of Dairy Science* **87**: 757-763.