

New Zealand Society of Animal Production online archive

This paper is from the New Zealand Society for Animal Production online archive. NZSAP holds a regular annual conference in June or July each year for the presentation of technical and applied topics in animal production. NZSAP plays an important role as a forum fostering research in all areas of animal production including production systems, nutrition, meat science, animal welfare, wool science, animal breeding and genetics.

An invitation is extended to all those involved in the field of animal production to apply for membership of the New Zealand Society of Animal Production at our website www.nzsap.org.nz

[View All Proceedings](#)

[Next Conference](#)

[Join NZSAP](#)

The New Zealand Society of Animal Production in publishing the conference proceedings is engaged in disseminating information, not rendering professional advice or services. The views expressed herein do not necessarily represent the views of the New Zealand Society of Animal Production and the New Zealand Society of Animal Production expressly disclaims any form of liability with respect to anything done or omitted to be done in reliance upon the contents of these proceedings.

This work is licensed under a [Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License](http://creativecommons.org/licenses/by-nc-nd/4.0/).



You are free to:

Share— copy and redistribute the material in any medium or format

Under the following terms:

Attribution — You must give [appropriate credit](#), provide a link to the license, and [indicate if changes were made](#). You may do so in any reasonable manner, but not in any way that suggests the licensor endorses you or your use.

NonCommercial — You may not use the material for [commercial purposes](#).

NoDerivatives — If you [remix, transform, or build upon](#) the material, you may not distribute the modified material.

<http://creativecommons.org.nz/licences/licences-explained/>

BRIEF COMMUNICATION

Efficacy of antibiotic treatment at drying off in curing existing infections and preventing new infections in dairy goats

F.M. ANNISS AND S. MCDOUGALL

Animal Health Centre, PO Box 21, Morrinsville, New Zealand.

Infusion of antibiotics into the udder at the end of the lactating period ("dry cow therapy"; DCT) is an essential control measure for bovine mastitis (Smith *et al.*, 1967). Dry cow therapy eliminates existing intramammary infections (IMI) and reduces the incidence of new infections over the dry period with a resultant reduced prevalence of IMI at the commencement of the subsequent lactation (Smith *et al.*, 1967; Harmon *et al.*, 1986). The cure rate of existing infections after DCT has been reported as between 60-90% (Pankey *et al.*, 1982; Sol *et al.*, 1994; Browning *et al.*, 1990; Williamson *et al.*, 1995). Dry cow therapy reduces the new IMI rate by approximately 50% (Browning *et al.*, 1994; Williamson *et al.*, 1995).

Infusion of antibiotics at the end of lactation in dairy goats ("dry-goat therapy"; DGT) has been reported to cure between 66% and 100% of existing IMI (Fox *et al.*, 1992; Poutrel & de Cremoux, 1995; Poutrel *et al.*, 1997). However, the efficacy of DGT in reducing the new infection rate over the dry period has not been examined.

The aim of this trial was to assess the efficacy of intramammary antibiotic treatment of dairy goats at the end of lactation in curing existing mammary gland infections and reducing the incidence of new infections over the dry period and early in the subsequent lactation.

Duplicate milk samples were taken from 476 dairy goats from four herds, seven days prior to the end of lactation. Milk samples of ~5 ml were obtained from each gland following aseptic preparation of the teat end by scrubbing with a cotton wool ball moistened in 70% methylated spirits and cultured for bacteria. A single ~20 ml sample was also taken and analysed for somatic cell count. Animals were blocked by bacteriological status (0 = uninfected, 1 = one gland infected, 2 = two glands infected), ranked on age (1, 2, >3 years) and mean SCC of both glands seven days before the end of lactation, then randomly assigned to one of two groups within sequential pairs of goats within herds. Following the final milking, each gland within the treatment group was aseptically prepared as above and infused with a single intramammary syringe containing 300 mg of procaine penicillin, 100 mg of dihydrostreptomycin and 100 mg of nafcillin (Norfpenzal D.C., Intervet, Auckland, New Zealand) while the other does were left as untreated controls. Antiseptic (0.5% iodine in H₂O) was applied to the teat end by spraying within 1 minute of treatment to all goats. Single milk samples were collected before the final milking for bacteriology. Duplicate milk samples were collected between 0 and 4 days post-kidding and again 3 to 4 days later for bacteriology.

For bacterial culture, a sub-sample of milk (10 µl)

was streaked onto Columbia Sheep Blood (5%) Agar (Forte Richard Laboratories, Auckland) and incubated for 48 hours at 37°C. Bacteria were speciated on the basis of colony morphology, Gram stain reaction, catalase, coagulase and CAMP tests, using standard procedures from the National Mastitis Council Laboratory Handbook on Bovine Mastitis (Anon, 1999).

A gland was defined as infected if culture of two of the three pre-dry off samples resulted in growth of three or more colonies of the same bacterial species. A gland was defined as cured if the bacterial species present at drying off was not isolated from any of the samples taken post kidding. A gland was defined as newly infected if no bacteria were isolated at drying off and then three or more colonies of the same bacteria were present in either of the post-kidding samples, or if the bacterial species isolated post-kidding was different from the species isolated pre-dry off.

Data were analysed at gland level and it was assumed that glands within a goat were independent. Backward stepwise logistic regression was used to analyse cure and new infection rates. Initial analysis of the main effects (age, herd, treatment, bacterial species at drying off and length of the non-lactation period) was performed using univariate analysis (either χ^2 or logistic regression) and factors found to be associated ($P < 0.2$) were then offered to the backward stepwise models. A number of glands were missed or the samples contaminated at subsequent samplings and were removed from the analyses.

The prevalence of IMI before drying off was 27.2% (258/948) of glands. Pathogens isolated included coagulase negative staphylococcus (CNS) (59.3% of all IMI), *Corynebacterium sp.* (Coryn) (29.1%), *Staphylococcus aureus* (SA) (6.2%), *Streptococcus agalactiae* (SAG) (4.3%) and other species (1.2%). Cure rate increased following treatment ratio (OR = 46.6 for treated compared to control; $P < 0.01$; Table 1), decreased with age (38/56 (67.9%) vs. 97/167 (58.1%), OR = 0.46, $P = 0.07$) for 1-year-olds compared to > 1-year-olds, respectively) and was lower for SA infections than other bacterial species (OR = 0.74, 0.06, 2.34, $P = 0.50$, 0.00, 0.39 for Coryn, SA and SAG, respectively; with CNS as the reference category).

Treatment reduced the likelihood of new IMI compared to untreated controls (OR = 0.14, $P < 0.01$; Table 1). Older animals were more likely to acquire a new IMI than younger animals (8/287 (2.8%), 12/245 (4.9%) and 29/314 (9.2%) for 1, 2 and >2-year-olds respectively, $P < 0.01$). Glands with an IMI at dry off were less likely to acquire a new IMI than glands uninfected at dry off (6/238 (2.5%) vs. 43/609 (7.1%), OR = 0.15, $P < 0.01$). There

TABLE 1: Summary of numbers and percentages of glands infected or uninfected at drying off and following kidding for goats treated by intramammary infusion of antibiotics (1 syringe of penicillin, dihydrostreptomycin and nafcillin per gland) at drying-off (treated) or left as untreated controls (Control).

Status at end of lactation		Enrolled n		Cured		New infections		
				Total	%	n	Total	%
Uninfected	Treated	311				6	311	1.9
	Control	298				37	298	12.4
	Total	609				43	609	7.1
Infected	Treated	114	99	107	92.5	3	113	2.7
	Control	129	36	116	31.0	3	125	2.4
	Total	243	135	223	60.5	6	238	2.5

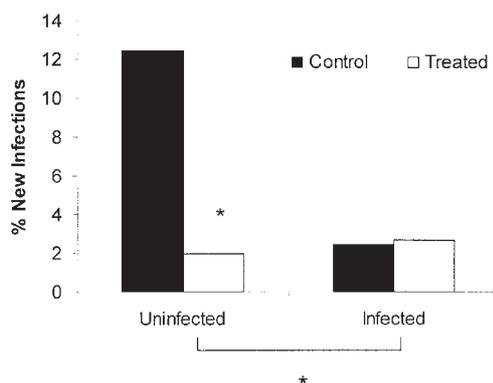
was a treatment by infection status interaction ($P < 0.05$, Figure 1) as control glands within the uninfected group had a higher new IMI than treated but uninfected glands. Among the infected glands treatment had no effect on new IMI rate.

This trial demonstrated that infusion of antibiotics at drying off into the goat mammary gland reduced the number of new infections over the dry period and increased the cure rate of existing infections.

The cure rate of existing infections was 92.5% in treated and 31% in untreated glands which is similar to previous reports for goats (Fox *et al.*, 1992; Poutrel & de Cremoux, 1995) and cattle (Smith *et al.*, 1967; Sol *et al.*, 1994; Williamson *et al.*, 1995). In common with reports for cattle, the SA cure rates were low. This has been associated with poor penetration and distribution of antibiotics throughout the gland (Owens & Nickerson, 1990; Sol *et al.*, 1994; Sol *et al.*, 1997; Owens *et al.*, 1997) and the ability of SA to survive intracellularly (Owens & Nickerson, 1990). Similar to cattle, the higher cure rate in the younger animals may be due to a shorter duration of infection before treatment or reduced immunocompetence with age (Sol *et al.*, 1994; Sol *et al.*, 1997).

The number of new IMI across the non-lactating period was reduced from 9% to 2% following DGT which

FIGURE 1. New intramammary infection rate (% glands newly infected) over the non-lactation period in glands from dairy goats defined as uninfected or infected before drying-off and treated or not treated at the end of lactation with intramammary antibiotics.



is consistent with cattle studies (Smith *et al.*, 1966; Browning *et al.*, 1990; Williamson *et al.*, 1995). New IMI rate was reduced by the presence of an IMI at drying off, and has been reported in dairy cattle, however the mechanism for this is not clear (Pankey *et al.*, 1982; Pankey *et al.*, 1985; Rainard & Poutrel, 1988).

It is concluded that infusion of antibiotics into the mammary gland of dairy goats at the end of lactation results in an increased cure rate of existing infections and a reduction in new infection rate. Hence dry-goat therapy is a useful tool for managing milk quality in dairy goats.

ACKNOWLEDGEMENTS

The technical assistance of Helena Habgood is gratefully acknowledged. Financial support was provided by a Foundation for Research, Science and Technology grant (AHC 001) and a Barbara Smith Scholarship from the New Zealand Veterinary Association. The antibiotics were kindly provided by Intervet New Zealand limited. The cooperation of the herdowners is gratefully acknowledged.

REFERENCES

- Anon; 1999: Laboratory Handbook on Bovine Mastitis, National Mastitis Council, Madison, Wisconsin, USA.
- Browning, J.W.; Mein, G.A.; Barton, M.; Nicholls, T.J.; Brightling, P. 1990: Effects of antibiotic therapy at drying off on mastitis in the dry period and early lactation. *Australian veterinary journal* 67: 440-442.
- Browning, J.W.; Mein, G.A.; Brightling, P.; Nicholls, T.J.; Barton, M. 1991: Strategies for mastitis control - dry cow therapy and culling. *Australian veterinary journal* 71: 179-181.
- Fox, L.K.; Hancock, D.D.; Horner, S.D. 1992: Selective intramammary antibiotic therapy during the non-lactating period in goats. *Small ruminant research* 9: 313-318.
- Harmon, R.J.; Crist, W.L.; Hemken, R.W.; Langlois, B.E. 1986: Prevalence of minor udder pathogens after intramammary dry treatment. *Journal of dairy science* 69: 843-849.
- Owens, W.E.; Nickerson, S.C. 1990: Treatment of *Staphylococcus aureus* with penicillin and novobiocin: Antibiotic concentrations and bacteriological status in milk and mammary tissue. *Journal of dairy science* 73: 115-124.
- Owens, W.E.; Ray, C.H.; Watts, J.L.; Yancey, R.J. 1997: Comparison of success of antibiotic therapy during lactation and results of antimicrobial susceptibility tests for bovine mastitis. *Journal of dairy science* 80: 313-317.
- Pankey, J.W.; Barker, R.M.; Twomey, A.; Duirs, G. 1982: A note on the effectiveness of dry cow therapy in New Zealand dairy herds. *New Zealand veterinary journal* 30: 50-52.
- Pankey, J.W.; Nickerson, S.C.; Boddie, R.L.; Hogan, J.S. 1985: Effects of *Corynebacterium bovis* infection on susceptibility to major mastitis pathogens. *Journal of dairy science* 68: 2684-2693.
- Poutrel, B.; de Cremoux, R. 1995: Efficacy of antibiotic treatments at drying-off for udder infections in goats. In: Saran, A.; Soback, S. *ed.* Proceedings of the Third IDF International Mastitis Seminar, Tel Aviv, Israel, pp 91-92.
- Poutrel, B.; de Cremoux, R.; Ducelliez, M.; Verneau, D. 1997: Control of intramammary infections in goats - impact on somatic cell counts. *Journal of animal science* 75: 566-570.
- Rainard, P.; Poutrel, B. 1988: Effect of naturally occurring intramammary infections by minor pathogens on new infections by major pathogens in cattle. *American journal of veterinary research* 49: 327-329.
- Smith, A.; Neave, F.K.; Dodd, F.H. 1966: Methods of reducing the incidence of udder infection in dry cows. *Veterinary record* 79: 233-235.
- Smith, A.; Westgarth, D.R.; Jones, M.R.; Neave, F.K.; Dodd, F.H.; Brander, G.C. 1967: Methods of reducing the incidence of udder infection in dry cows. *Veterinary record* 81: 504-510.

- Sol, J.; Sampimon, O.C.; Snoep, J.J.; Schukken, Y.H. 1994: Factors associated with bacteriological cure after dry cow treatment of subclinical staphylococcal mastitis with antibiotics. *Journal of dairy science* 77: 75-79.
- Sol, J.; Sampimon, O.C.; Snoep, J.J.; Schukken, Y.H. 1997: Factors associated with bacteriological cure during lactation after therapy for subclinical mastitis caused by *Staphylococcus aureus*. *Journal of dairy science* 80: 2803-2808.
- Williamson, J.H.; Woolford, M.W.; Day, A.M. 1995: The prophylactic effect of a dry-cow antibiotic against *Streptococcus uberis*. *New Zealand veterinary journal* 43: 228-234.