

New Zealand Society of Animal Production online archive

This paper is from the New Zealand Society for Animal Production online archive. NZSAP holds a regular annual conference in June or July each year for the presentation of technical and applied topics in animal production. NZSAP plays an important role as a forum fostering research in all areas of animal production including production systems, nutrition, meat science, animal welfare, wool science, animal breeding and genetics.

An invitation is extended to all those involved in the field of animal production to apply for membership of the New Zealand Society of Animal Production at our website www.nzsap.org.nz

[View All Proceedings](#)

[Next Conference](#)

[Join NZSAP](#)

The New Zealand Society of Animal Production in publishing the conference proceedings is engaged in disseminating information, not rendering professional advice or services. The views expressed herein do not necessarily represent the views of the New Zealand Society of Animal Production and the New Zealand Society of Animal Production expressly disclaims any form of liability with respect to anything done or omitted to be done in reliance upon the contents of these proceedings.

This work is licensed under a [Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License](http://creativecommons.org/licenses/by-nc-nd/4.0/).



You are free to:

Share— copy and redistribute the material in any medium or format

Under the following terms:

Attribution — You must give [appropriate credit](#), provide a link to the license, and [indicate if changes were made](#). You may do so in any reasonable manner, but not in any way that suggests the licensor endorses you or your use.

NonCommercial — You may not use the material for [commercial purposes](#).

NoDerivatives — If you [remix, transform, or build upon](#) the material, you may not distribute the modified material.

<http://creativecommons.org.nz/licences/licences-explained/>

BRIEF COMMUNICATION

Susceptibility of the breeding ewe to parasitism

R.W. McANULTY, A.S. FAMILTON AND A.R. SYKES

Animal and Veterinary Sciences Group, Lincoln University, Canterbury, New Zealand.

ABSTRACT

Pregnant Coopworth ewes were allocated to four periods of parasitic challenge which commenced either -4, 0, 6 or 12 weeks from lambing. Within each period, ewes were further allocated to four groups (n=6) viz initial slaughter; treatment with anthelmintic followed by a single infection with 20,000 larvae of *Ostertagia (Teladorsagia) circumcincta* and slaughtered after 21 days; chronic infection with 4000 larvae per day for 30 days or no infection. Both these latter groups were again dosed with anthelmintic on day 50 and given 20,000 larvae on day 57 and slaughtered 21 days later. At the start of each period 6 non-breeding ewes were challenged with 20,000 larvae and slaughtered 21 days later. All ewes were housed indoors. Significantly higher worm burdens were found at all times in response to challenge in breeding ewes compared to non-breeding ewes, except during period 4 (12 weeks after lambing) where differences were non-significant. Host suppression of faecal egg output was seen during periods 2 and 3, with egg output being reduced by 50% and 95% respectively, compared to that seen during period 1. Faecal egg counts and worm burdens of previously infected ewes were lower than their initially non-exposed counterparts ($P < 0.01$). Even though immunity was relaxed during late pregnancy and early lactation exposure to infection was nevertheless important in stimulating the re-establishment of immune competency.

Keywords *Ostertagia*, artificial, challenge, parturition, breeding ewes, nematode, parasites, susceptibility.

INTRODUCTION

The occurrence of a peri-parturient rise in faecal egg output is well documented (Reid and Armour, 1975b; O'Sullivan and Donald, 1973). The duration of this period of susceptibility to parasitic infection in ewes has not yet been clearly defined. This is critical in determining not only the extent of production losses which occur in the ewes (Leyva *et al.*, 1982; Thomas and Ali, 1983; Sykes and Juma, 1984), but also in determining when and whether the ewes should be given anthelmintics to reduce subsequent pasture larval contamination and production loss. The resistance of the ewe to re-infection following anthelmintic treatment during lactation has yet to be assessed.

This study compares the response of breeding and non-breeding ewes to artificial challenge at intervals from late pregnancy until late lactation and also examines the effect of prior larval exposure on the ewes' response to re-infection.

MATERIALS AND METHODS

Five-year old pregnant twin bearing and non-pregnant Coopworth ewes were allocated to four experimental periods in which challenge commenced either -4, 0, 6 or 12 weeks from lambing. Within each period, ewes were further allocated to four groups (n=6) viz initial slaughter (ISG); or treatment with anthelmintic (Ivermectin MSD Agvet NZ Ltd, 200 mcg/kg bodyweight) followed by a single infection with 20,000 larvae of *Ostertagia (Teladorsagia) circumcincta* and slaughtered after 21 days (SAI); chronic infection with 4000 larvae per day for 30 days (MCI) or no infection (NIC). Both these latter groups were again dosed with anthelmintic on day 50 and given a 20,000 larvae challenge on day 57 and slaughtered 21 days later. In each period non-breeding ewes (NBSAI) were treated as for group SAI. All ewes were housed indoors and offered a pelleted diet. Faecal egg counts were measured weekly and worm burdens determined at slaughter.

RESULTS

TABLE 1 Mean worm burden in breeding (SAI) and non-breeding (NBSAI) ewes 21 days after challenge with 20,000 *Ostertagia circumcincta* larvae on entry to the experiment.

Time of challenge Period	Wks before or after lambing	Mean total worm burden	
		SAI ewes	NBSAI ewes
1	-4 wk	1194**	224
2	0 wk	2564**	348
3	6 wk	1877**	385
4	12 wk	222	220

** P<0.01

Significantly higher worm burdens (P<0.01) were found at all times in response to challenge in breeding ewes (SAI) compared to non-breeding ewes (NBSAI), except during period four (Table 1).

TABLE 2 Effect of recent chronic exposure (MCI-4000 *Ostertagia circumcincta* larvae/d for 30 days) or non exposure (NIC) on the worm burdens of breeding ewes after challenge with 20,000 *Ostertagia circumcincta* larvae.

Time of challenge Period	Wks after lambing	Total worm burden	
		MCI ewes	NIC ewes
1	4-7	473	2040**
2	8-11	213	681**
3	14-17	22	25
4	20-23	275	56

** P<0.01

Maximum worm burdens occurred around parturition but by 12 weeks after lambing worm burdens in breeding ewes were similar to those found in non-breeding ewes throughout the experiment. The magnitude and duration of the rise in the mean egg production of MCI ewes, in response to a trickle challenge, decreased from period 1 to 4. Faecal egg counts of MCI ewes peaked 6 weeks after infection commenced, with maximum levels of 1330, 380, 80, and 16 epg recorded for periods 1 to 4 respectively. The

time required for the infection to develop to patency, as gauged by faecal egg counts, increased during the course of the trial. Nematode egg production in MCI ewes was first recorded 14, 21, 35 and 42 day after infection commenced, for periods 1 to 4 respectively.

Both the worm burdens and faecal egg counts of previously infected ewes (MCI) were lower than their initially non-exposed counterparts (NIC) in response to challenge on day 57 during periods 1 and 2 (P<0.01; Table 2).

DISCUSSION

The worm burden which becomes established is a function of the larval challenge and the change in the susceptibility of the host. Non-breeding ewes clearly showed that the challenge experienced was within their capabilities to resist. On the other hand breeding ewes suffered loss of resistance prior to, as well as after parturition as judged by establishment of nematode populations in the field and as a result of artificial challenge, confirming observations of Connan (1968) and Reid and Armour (1975a, 1975b). Maximum susceptibility appeared to be around or soon after parturition but a reduced immune response was still apparent 6 weeks after parturition, confirming previous suggestions of Reid and Armour (1975a), Leyva *et al.* (1982), Sykes and Juma (1983), and Thomas and Ali (1984), though by this stage host resistance to infection in this study was increasing.

The time required for patency of the infection to develop, as gauged by faecal egg count data from periods 2, 3 and 4, increased over the experimental periods from 3 to 7+ weeks and the decline in faecal egg output in response to trickle infections, with time indicates increasing host resistance to parasitic challenge occurring after or about 6 weeks of lactation. Gibbs and Barger (1986) reported a similar suppression of egg output in lactating ewes despite worm burdens remaining constant.

Faecal egg counts (period 1) and worm burdens (period 1 and 2) of previously infected ewes (MCI) were lower than their non-exposed counterparts (NIC) after challenge on day 57, indicating that larval stimulus during late pregnancy and early lactation evoked a substantial immune response. Lactating ewes, with recent larval exposure during pregnancy, were

found by Leyva *et al.* (1982) to be more resistant as judged by faecal egg count, food intake response and bodyweight than their non-exposed counterparts. Re-establishment of resistance after its loss around parturition may well be dependant on continuing larval challenge.

REFERENCES

- Connan, R.M. (1968a) Studies on the worm populations in the alimentary tract of breeding ewes. *Journal of Helminthology* 42: 9-28.
- Gibbs, H.C.; Barger, I.A. (1986) *Haemonchus contortus* and other trichostrongylid infections in parturient, lactating and dry ewes. *Veterinary Parasitology* 22: 57-66.
- Leyva, V.; Henderson, A.E.; Sykes, A.R. (1982) Effect of daily infection with *Ostertagia circumcincta* larvae on food intake, milk production, and wool growth in sheep. *Journal of Agriculture Science, Cambridge* 99: 249-259.
- O'Sullivan, B.M.; Donald, A.D. (1973) Response to infection with *Haemonchus contortus* and *Trichostrongylus colubriformis* in ewes of different reproductive status. *International Journal for Parasitology* 3: 521-530.
- Reid, J.F.S.; Armour, J. (1975a) Studies in Scottish hill sheep. *Journal of Comparative Pathology* 85: 163-170.
- Reid, J.F.S.; Armour, J. (1975b) Seasonal variations in the gastrointestinal nematode populations of Scottish hill sheep. *Research in Veterinary Science* 18: 303-313.
- Sykes, A.R.; Juma, M.H. (1984) Effect of chronic experimental infection with *Ostertagia circumcincta* and anthelmintic therapy on the performance of lactating sheep at pasture. *New Zealand Journal of Experimental Agriculture* 12: 243-249.
- Thomas, R.J.; Ali, D.A. (1983) The effect of *Haemonchus contortus* infection on the pregnant and lactating ewe. *International Journal for Parasitology* 13: 393-398.